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Targeted Drug Delivery to The Brain for Treatment of Nipah Encephalitis

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Abstract: Nipah virus (NiV), which is a henipavirus, is a contagious and lethal illness with substantial morbidity as a result of its association with severe acute respiratory disease and encephalitis, and a high fatality rate. There are currently no antiviral agents available for treating the neurological disease produced by Nipah virus infection despite the potential for outbreaks throughout the world. One of the main reasons for this lack of treatment for central nervous system infections is the presence of the blood-brain barrier (BBB). Because the BBB restricts drug movement from circulation into the brain, it limits the availability of drugs for treating CNS infections. There have been several advances in developing targeted drug delivery for CNS infections and these new methods of delivering drugs have demonstrated the ability to overcome this barrier. There has been active research into developing nanocarrier systems to carry drugs across the BBB that will improve the efficacy of the treatment of infections in the CNS, including various types of liposomes, polymeric nanoparticles, solid lipid nanoparticles, dendrimers, and nanogels. There has also been research into intranasal (nasal) delivery methods and receptor-mediated strategies as potential methods of delivering drugs directly to the site of infective agents in the brain through the nasal mucosa. The use of these strategies may improve the effectiveness of delivering antivirals, monoclonal antibodies, and nucleic acid based therapeutics to infected tissues in the brain while reducing the potential for adverse effects in other parts of the body. This review article discusses the pathophysiology of Nipah Encephalitis, the challenges to traditional methods for treating the viral infection, as well as more recent advances in systems designed for targeted delivery of drugs directly to the site of infection of the CNS. The review will also provide a summary of common themes in current research on development of new delivery SYSTEMS for developing new antivirals. Issues of limitations and potential areas of future research development will also be addressed within the context of this review article.

keywords: Nipah Virus (NiV), Nipah Encephalitis ,Targeted Drug Delivery ,Blood-Brain Barrier (BBB) ,Brain Drug Delivery ,Nanoparticles, Liposomes, Intranasal Delivery, Antiviral Therapy, Nanomedicine, Central Nervous System (CNS), Receptor-Mediated Transport

INTRODUCTION

Overview of Nipah Virus

The only bats that can transmit the Nipah virus to people are fruit bats; therefore, it is very serious and can be transmitted to both people and animals (It may cause death). When some pig farmers contracted the

virus in Malaysia (1998-1999), the Nipah virus was first identified in people. People can become infected with the Nipah virus by coming into contact with infected fruit bats' saliva or urine, touching infected animals, consuming food that has been tainted by infected fruit bat saliva or urine, or touching the blood or saliva or urine of [an infected person]. Many signs are associated with Nipah virus in humans; some examples are minor fever, severe breathing difficulties and neurological conditions such as [encephalitis].

There have been multiple Nipah virus outbreaks through various locations in both South and Southeast Asia, such as Bangladesh and India. The case fatality rate associated with these outbreaks is reported to be between 40%-75% depending on the conditions of each particular outbreak, healthcare infrastructure and severity of disease during each occurrence of the outbreak. The human-to-human transmission of the Nipah virus can lead to severe neurological problems, creating a major public health crisis. There are no registered vaccines and there are no antivirals for treating Nipah virus, thus adding to the potential threat of future outbreaks. Nipah virus has therefore been categorised as a priority pathogen by international health bodies and further intensive research into and development of effective preventative and therapeutic measures is urgently needed.

Infections with the Nipah virus can cause encephalitis, which may be extremely severe and substantially decrease the quality of life for survivors. In addition to causing encephalitis, Nipah virus infections can also invade and inflame the neurons and central nervous system (CNS) directly, thereby damaging the CNS and neuronal tissues. The presence of the blood-brain barrier (BBB) limits the use of drug therapies that could otherwise help heal the CNS and brain tissue. Researchers are now developing targeted drug-delivery systems that are capable of crossing or circumventing the effects of the BBB. Promising advanced drug delivery systems used in the prevention and treatment of Nipah encephalitis include nanoparticle-based carriers (e.g., liposomal formulations); receptor-mediated targeting; and intranasal delivery.¹⁻⁶

NIPAH VIRUS AND ENCEPHALITIS

Virology of Nipah Virus

Nipah Virus belongs to the Henipaviruses genus and is part of the Paramyxoviridae family. The genetic material for this virus is RNA. The total number of bases in Nipah Virus' nucleotide sequence is an approximate number of 18000 bases, which is made up of 18358 bases total. The virion/ particle structure contains six structural proteins that perform unique functions related to the structure. There are 6 proteins associated with Nipah Virus: 1. Nucleoprotein. 2. Phosphoprotein 3. Matrix protein (M), 4. F Protein (F), 5. Glycoprotein (G), 6. Large Polyprotein (L) The virion attaches to host cell receptors by way of G and F glycoproteins binding to ephrin receptors B2 and B3 on the basolateral membrane of endothelial and neuronal type cells, respectively, and the sites for binding are found in the host cell cytoplasm (intracellular). Due to the large number of these receptors expressed on the surface of these tissues, nipah virus has the ability to be disseminated throughout all regions of the host body including the Central Nervous System (CNS). The sole natural host for nipah viruses are the fruit bats, generally classified as (Genus: Pteropus). Humans can be infected via contact with infected animals, eating or drinking contaminated food products, or via contact with an infected individual.

Pathogenesis of Nipah Encephalitis

The breakdown of the body's multiple cell types occurs as the Nipah virus replicates within different body tissues. As the viral infection destroys cells, it eventually progresses to infect the CNS and PNS systems (central nervous system and peripheral nervous system). After the virus has been introduced into the body through the respiratory system, it begins to infect epithelial cells in the airway and adjacent peribronchial lymph nodes. From these sites, the virus can spread to other tissues in the body through the blood. The Nipah virus infects endothelial cell populations (which make up the inner lining of blood vessels) leading to the development of vasculitis, destruction of endothelial cells (including the thinning of the endothelium), thrombosis, and disruption of the blood-brain barrier. These changes create a pathway to enter the brain where it infects/inflames the neurons and causes extensive neuronal damage. The immune reaction to the virus through the release of pro-inflammatory cytokines and chemokines contributes further injury to the neurons. The most common histopathological features of affected brains include vasculitis, microinfarcts, degeneration of neurons, and widespread signs of encephalitis, all contributing to the neurological manifestations of Nipah virus.

Clinical manifestation

Nipah virus infections have an incubation period that often falls between 4-14 days, although there have been some reports of longer incubation periods. The clinical symptoms range from Asymptomatic to clinically severe and fatal disease.

The early symptoms of Nipah virus infections are typically as follows:

1. Fever
2. Headache
3. Myalgias (muscle aches)
4. Fatigue
5. Sore throat
6. Dizziness

The further the patient progresses with infection, patients may experience respiratory signs including:

1. Cough
2. Shortness of Breath
3. Acute Respiratory Distress

Neurologically, patients may present with signs of:

1. Altered Mental Status
2. Confusion
3. Drowsiness
4. Seizures
5. Encephalitis
6. Coma

Patients who suffer from serious illness usually have fast decline in their neurological condition within a couple of days before they develop respiratory failure and die; if the patient survives, he/she may develop long-term problems with cognition (thinking), personality, weakness, and later on develop encephalitic symptoms from Nipah virus infection. There are a high mortality rate due to Nipah virus and a high rate of neurological problems with this infection which highlights the immediate need for an effective way to treat patients. This creates a need for targeted delivery methods to bring drugs to the brain in order to treat patients effectively. Nipah virus infections have an incubation period that often falls between 4-14 days, although there have been some reports of longer incubation periods. The clinical symptoms range from Asymptomatic to clinically severe and fatal disease.

CHALLENGES IN TREATING NIPAH ENCEPHALITIS

There are numerous challenges in treating Nipah encephalitis, due mostly to the blood–brain barrier (BBB), a physiological barrier that separates the blood that circulates in the body from the central nervous system (CNS). The BBB is an extremely selective barrier made up mostly of endothelial cell junctions and astrocytes, however there are also some pericytes and basement membrane components. The BBB plays an essential role in regulating the passage of substances, specifically the passage of molecules into the CNS from the bloodstream; and protecting the CNS from toxins, pathogens and other harmful agents.

Even though the BBB helps to protect and maintain the homeostasis of the brain, because of its structure (endothelial cells tightly bound together), it prevents many therapeutic drugs (antiviral drugs, monoclonal antibodies, and large biomolecules) from entering the CNS. Therefore, the conventional delivery of drugs into the CNS typically results in the inadequate concentrations of the drugs, at the site of infection because the Nipah virus can invade and replicate within nervous tissue. As a result, developing targeted drug delivery systems that can cross the BBB or bypass it is Important in the development of effective therapies for Nipah encephalitis. There are currently no specific antiviral therapies approved for

the treatment of Nipah virus infection. Conventional therapy is currently limited to supportive care, including providing respiratory support and the providing fluids.³⁻⁶

BLOOD BRAIN BARRIER

Blood–Brain Barrier (BBB): Structure and Components

The blood–brain barrier (BBB) is a highly specialized and dynamic interface that separates the circulating blood from the central nervous system (CNS). Its primary function is to maintain cerebral homeostasis by regulating the movement of ions, nutrients, metabolites, and immune cells between the bloodstream and the brain. While this selective permeability is essential for protecting neural tissue from toxins and pathogens, it also presents a major obstacle to the delivery of therapeutic agents for neurological disorders, including Nipah virus (NiV) encephalitis. The BBB is formed by a coordinated network of cellular and extracellular components collectively referred to as the neurovascular unit (NVU). The NVU comprises brain microvascular endothelial cells, tight junction proteins, pericytes, astrocytes, the basement membrane, neurons, and resident immune cells. These components work together to preserve barrier integrity, regulate cerebral blood flow, and control molecular transport into the CNS.

Brain Microvascular Endothelial Cells

Brain microvascular endothelial cells (BMECs) constitute the structural foundation of the BBB. Unlike endothelial cells in peripheral tissues, BMECs possess continuous, non-fenestrated membranes and exhibit extremely low rates of pinocytosis. They contain numerous mitochondria to support the energy-dependent transport systems required for maintaining BBB function. These endothelial cells express specialized transport proteins that regulate the influx of essential nutrients such as glucose, amino acids, vitamins, and ions while simultaneously preventing the entry of potentially harmful substances. Efflux transporters, including P-glycoprotein (P-gp), breast cancer resistance protein (BCRP), and members of the multidrug resistance-associated protein (MRP) family, actively remove xenobiotics and many therapeutic drugs from the brain, thereby limiting drug accumulation within the CNS. During Nipah virus infection, inflammatory mediators released in response to viral replication can alter endothelial cell function, increasing vascular permeability and contributing to neurological complications.

Tight Junction Proteins

Adjacent endothelial cells are interconnected by tight junction complexes, which are responsible for the exceptionally low paracellular permeability of the BBB. These complexes are composed primarily of transmembrane proteins such as claudin-5, occludin, and junctional adhesion molecules (JAMs), together with cytoplasmic scaffold proteins including zonula occludens-1 (ZO-1), ZO-2, and ZO-3. Tight junction proteins restrict the passive diffusion of hydrophilic molecules and pathogens into the brain, forcing most substances to cross the BBB through tightly regulated transcellular transport mechanisms. Experimental studies have demonstrated that viral infections and neuroinflammatory responses can disrupt tight junction integrity through cytokine-mediated signaling pathways, resulting in increased BBB permeability. In Nipah virus encephalitis, impairment of tight junctions may facilitate immune cell infiltration into the CNS while also contributing to cerebral edema and neuronal injury.

Pericytes

Pericytes are contractile mural cells embedded within the basement membrane and closely associated with brain capillary endothelial cells. They play an essential role in BBB formation, stabilization, and maintenance throughout life. Pericytes regulate endothelial cell proliferation, angiogenesis, capillary blood flow, and extracellular matrix production. They also influence tight junction assembly and suppress endothelial transcytosis, thereby contributing to the low permeability of the BBB. Loss or dysfunction of pericytes has been associated with increased vascular leakage, impaired cerebral perfusion, and enhanced neuroinflammation. During viral encephalitis, inflammatory signals may alter pericyte function, promoting BBB disruption and facilitating viral dissemination within the brain.

Astrocytes

Astrocytes are star-shaped glial cells whose terminal processes, known as astrocytic end-feet, envelop nearly the entire surface of brain capillaries. Although astrocytes do not directly form the BBB, they are

indispensable for its development and maintenance. Astrocytes secrete numerous signaling molecules, including transforming growth factor- β (TGF- β), glial-derived neurotrophic factors, and angiopoietins, which promote endothelial differentiation and enhance tight junction integrity. They also regulate water and ion homeostasis through membrane proteins such as aquaporin-4 (AQP4) and inwardly rectifying potassium channels. In the setting of Nipah virus infection, activated astrocytes contribute to the neuroinflammatory response by releasing cytokines and chemokines. Persistent astrocyte activation may exacerbate neuronal injury while simultaneously altering BBB permeability.

Basement Membrane

The basement membrane is a specialized extracellular matrix that surrounds endothelial cells and pericytes, providing structural support to the neurovascular unit. It consists primarily of collagen type IV, laminins, nidogens, fibronectin, and heparan sulfate proteoglycans. Beyond its mechanical role, the basement membrane regulates cell adhesion, migration, differentiation, and molecular signaling. It also functions as an additional physical barrier that restricts immune cell migration into the CNS. Inflammatory enzymes such as matrix metalloproteinases (MMPs), which are upregulated during viral infections, can degrade basement membrane components, weakening BBB integrity and promoting inflammatory cell infiltration.

Neurons

Neurons communicate closely with vascular cells through neurovascular signaling pathways that regulate cerebral blood flow and metabolic activity. Although neurons are not structural elements of the BBB, they influence BBB function indirectly by interacting with astrocytes and endothelial cells. Neuronal injury resulting from Nipah virus infection triggers the release of inflammatory mediators and danger-associated molecular patterns that contribute to BBB dysfunction and further neuroinflammation.

Microglia

Microglia are the resident macrophages of the central nervous system and serve as the first line of immune defense against invading pathogens. Under physiological conditions, they continuously survey the neural environment and eliminate cellular debris. Following Nipah virus infection, microglia become activated and release inflammatory cytokines, chemokines, nitric oxide, and reactive oxygen species. While these responses contribute to viral clearance, excessive or prolonged activation can damage surrounding neurons and compromise BBB integrity. Microglial activation is therefore considered a key contributor to the neuropathogenesis of viral encephalitis.

Clinical Relevance to Nipah Virus Therapy

The highly selective architecture of the BBB significantly limits the penetration of antiviral drugs into the CNS, posing a major challenge in the treatment of Nipah virus encephalitis. Although inflammation during infection may transiently increase BBB permeability, this effect is unpredictable and often insufficient to achieve therapeutic drug concentrations within infected brain tissues. Understanding the structural and functional components of the BBB has facilitated the development of targeted drug delivery strategies, including liposomes, polymeric nanoparticles, dendrimers, exosomes, and receptor-targeted nanocarriers. These systems are designed to exploit endogenous transport mechanisms or receptor-mediated transcytosis to enhance drug delivery across the BBB while minimizing systemic toxicity. Such approaches hold considerable promise for improving the efficacy of antiviral therapies against Nipah virus and other neurotropic viral infections.⁷⁻¹⁰

Functions of the BBB

Protects brain against toxins and pathogens.

Maintains appropriate internally (For Homeostasis).

Controls transport of nutrients and waste products

Controls Access of drugs to the CNS.

Factors Affecting Drug Transport through the BBB:

1) Molecular Size

Small Molecules (<400-500 Da) will Cross More Easily.</mark>

Harder for Larger molecules to Cross the BBB.

2) Lipid solubility

Lipophilic (lipid soluble) cross the BBB More Readily than Hydrophilic molecules.

3) Charge

Neutral molecules cross the BBB More Easily.

Highly Ionized molecules do not cross the BBB as well

TARGETED DRUG DELIVERY STRATEGIES FOR THE BRAIN

Active Targeting

Active targeting is an approach that uses receptors and ligands which interact with appreciable blood-brain barrier (BBB) endothelial cell receptors or transport systems to make that regional delivery of therapeutic agents easier due to better access and therefore improve the amount of drug/deposit in the tissue by feeding into the system from within and it usually accomplishes this efficiently and in the right location than passive targeting.

Common ligands that may be used for targeting are:

- (1) transferrin
- (2) lactoferrin
- (3) apolipoprotein
- (4) peptides
- (5) monoclonal antibodies
- (6) folic acid.

When a ligand/receptor interaction occurs, a ligand/molecular transport occurs and that then provides for a mechanism of molecular transport across that BBB through usual cellular endocytosis, interaction with the lipid bilayer and cross-through the endothelial cell. Targeting via Receptor-Mediated Transport (RMT) compared to passive targeting would provide an opportunity for a more efficacious method and a more targeted location to deliver therapeutic agents using an active transport mechanism.

Active targeting of Ribavirin, Favipiravir, Remdesivir, Monoclonal Antibodies and other RNA-based therapies to the brain tissue of patients with Nipah viral illness may be accomplished as there would be more opportunity for efficacy and an increased amount of rainbow agent delivered to the brain.

Receptor-mediated transport

RMT is one of the most studied routes for targeting CNS endpoints in distributing CNS-directed therapeutics. It takes advantage of receptor-based pathways that exist at the BBB endothelial cells for the transport of therapeutics across into the CNS via transcytotic pathways.

Mechanism for transport via RMT (Receptor-Mediated Transport):

- (1) Targeting ligand binds to the apropos receptor at the luminal surface of the BBB endothelial cells.
- (2) The receptor-ligand complex is then taken by endocytosis.
- (3). A therapeutic agent can use transcytosis to cross the endothelial cell via a vesicle in order to reach the abluminal side (CNS) of the BBB.^{9,10,18,19}

There are 2 Receptors that can be utilized to transport CNS-directed therapeutics across the BBB include:

TfR (Transferrin Receptor)

Transport Via Carrier Mediated Mechanisms

Carrier mediated transport

Transport by Carrier Mediated Mechanisms (CMT) uses the body's natural shipping channels (transporters) for transporting vital materials (like sugars, amino acids, vitamins or nucleotides) to get past the endothelial barrier that protects your central nervous system (CNS).

Transportation using CMT is achieved by altering the structure of the drug or nanoparticle carrier to resemble the structural properties of molecules that would be recognized by an existing natural or genetically engineered transport system. This allows these modified drugs or nanoparticle carriers to utilize the existing transporters to move from the bloodstream to the CNS.

Transport Systems

Example of the Glucose Transporter (GLUT1)

GLUT1 is well represented in the blood-brain barrier and is responsible for transporting glucose across the blood-brain barrier (BBB) and serves as a model for designing improved nanoparticles for reaching the CNS.

Large Neutral Amino Acid Transporter (LAT1)

LAT1 is a transporter for several neutral amino acids, including phenylalanine and leucine. The conjugation of drug molecules with amino-acid analogues will allow the agent to be transported into the CNS by LAT1 transporter-mediated transport.

Nucleoside Transporters

The nucleoside transporters are responsible for allowing nucleosides and their respective analogues to enter the CNS. A number of antiviral agents are structurally similar to nucleosides and may benefit from their ability to use nucleoside transporter-mediated transport.⁹⁻¹⁸

Advantages

Efficient transport

low immunogenicity

use of natural physiological pathways

potential for repeated dosing.

Application in Nipah Virus Encephalitis

Carrier-mediated transport strategies may have applications to improve the CNS delivery of antiviral nucleoside analogues and other therapeutic agents to infected CNS tissues. The potential benefits from using CMT in combination with nanotechnology-based delivery devices are also possible.

NANOTECHNOLOGY-BASED DRUG DELIVERY SYSTEMS

Nanogels

Nanogels are nanosized, 3D hydrogel particles made of cross-linked polymer networks that can take up and hold lots of water while delivering therapeutics.

Structure

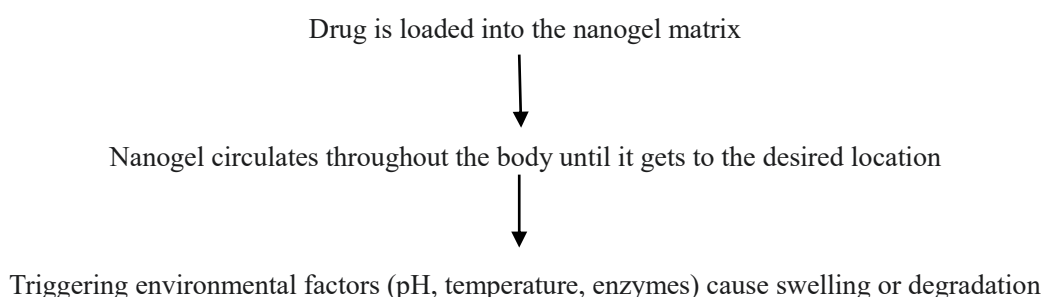
Made of hydrophilic polymers

Typically between 20-200nm

Can encapsulate drugs, proteins, peptides, and nucleic acids

Respond to your environment. That could mean responding to pH, temperature, and even enzymes.

Mechanism of Drug Delivery



Then, once it's at the tumor, the drug is slowly released so that just enough leaks out before all of the particles are broken down.

Advantages

High drug-loading capacity.

Excellent biocompatibility and biodegradability.

Controlled and sustained drug release.

Protection from degradation

Reduced toxicity and side effects

Ability to cross biological barriers such as the blood–brain barrier (BBB)

Applications

Brain-targeted drug delivery.

Cancer therapy.

Gene delivery.

Vaccine delivery Treatment of neurological disorders Infectious disease

Role in Nipah Encephalitis

Nanogels could help antiviral drugs get to the brain more effectively, boosting penetrance across the BBB and keeping release steady over time – making them a promising option for treating Nipah virus-triggered encephalitis^{11,12,16}

LIPOSOME-MEDIATED DRUG DELIVERY

The spherical characteristics of liposomes made them unique carriers for delivering drugs. Hydrophilic drugs can be delivered using liposomes, which are comprised of a phospholipid bilayer that encloses an aqueous compartment. These liposomes, along with being able to deliver hydrophobic drugs, can be created as drug delivery vehicles by incorporating various forms of lipophilic substances into their lipid bilayers and then delivering these substances to cells in the body through various methods. Because of this potential for use as a method of delivering various types of drug products (e.g., antiviral drugs, nucleotides, biologicals, etc.) to different organs in the body through a variety of methods, liposomal delivery systems have become a promising resource for drug formulation.

Typical liposomal products will include phospholipids as structural (i.e., phosphatidylcholine) components of the liposomal structure; cholesterol as a stabilizer for the liposome membrane structure; and surface modifier agents (e.g., PEG) to enhance circulation times (increased systemic half-lives) for therapeutic agents delivered to a patient. The incorporation of PEG as a surface modifier will increase the systemic half-life (stability) of the liposome and enhance liposome penetration into the brain by decreasing the amount of opsonization and that the mononuclear phagocytic system will uptake the liposome.

The enhancement of drug pharmacokinetics and pharmacodynamics by a liposome delivery system is one of the most significant advantages of using a liposomal delivery system. When a drug is encapsulated in a liposome, the drug in the liposomal formulation, has increased pharmacokinetic and pharmacodynamic activity.

Furthermore, liposomes have a biocompatible and biodegradable structure that provides less systemic toxicity than traditional dosage forms of medication.

With regard to the delivery of drugs into the CNS, it is possible to design liposomes as a delivery method.

Applications in Viral Encephalitis

Due to the significant consequences of severe encephalitis that causes extensive inflammation and neuronal dysfunction in the CNS and associated high mortality rates, CNS infections caused by viral

pathogens like Nipah virus provide a major challenge to traditional antiviral therapies as they are complicated by the presence of the BBB.

New methods of drug delivery using liposome-based formulations could potentially be used in overcoming the barriers presented by BBB. Numerous studies leveraging liposome-based drug delivery systems have been conducted where antiviral agents such as ribavirin, derivatives of acyclovir, analogues of oseltamivir, and nucleoside analogue compounds were loaded into the liposomes for evaluation of their efficacy against experimental models of CNS viral infections. The loading of antiviral agents into liposomes may provide added stability to the drug and slow the rate of release from the liposomal delivery system. The prolonged release of antiviral agents is particularly helpful for the management of rapidly progressive CNS viral infections. Investigation of liposomal formulations in experimental models of viral encephalitis demonstrated improved drug distribution to the brain and reduced systemic toxicity. Following the development of an inflamed state, tight junctions of endothelial cells in the BBB become disrupted leading to increased vascular permeability which enhances the ability of liposomes to cross or circumvent the BBB. This enhanced permeability and retention effect of inflamed tissues may further enhance the bulk accumulation of liposomal formulations in regions of the brain infected with the virus.

Currently, targeted liposomal formulations being developed for gene-related therapeutics include the delivery of small interfering RNA (siRNA) and messenger RNA (mRNA) that can be used to inhibit viral replication. With no approved medications for treating Nipah virus, the use of these new types of liposomal formulations will be important in the development of effective anti-Nipah virus therapies and for administering potential experimental antiviral products in the treatment of Nipah virus infections.¹³⁻¹⁶

RECENT ADVANCES AND CURRENT RESEARCH

A lot of recent research and progress on improving targeted drug delivery to the brain has been through developing nanoparticle based methods to cross the blood–brain barrier (BBB) for delivering drugs that will be delivered to brain tissue. There also has been some research completed with using receptor targeting, magnetic nanoparticles and using the intranasal route for drug delivery in order to improve therapeutic efficacy while minimizing the possibility of systemic side effects associated with the treatment of neurological infections, like Nipah encephalitis.

Pre-Clinical Research Studies

Pre-clinical studies based on the use of animal models have provided scientific evidence demonstrating the efficacy of a variety of nanotechnology based drug delivery systems. For instance, polymeric nanoparticles and lipid-based carriers have been shown to deliver effective antiviral and neuroprotective drugs across the BBB. Additionally, the use of intranasal formulations in delivery systems for medications allows for increased drug concentrations to be delivered to brain tissue through the direct nose-to-brain pathway, thus enhancing the bioavailability, localized action, and lowering toxicity when compared with traditional therapies.

Experimental Systems for Brain Targeting

The following experimental systems are currently being evaluated based on some research:

1. Receptor-mediated transport of nanoparticles that have been conjugated to ligands that would promote receptor-mediated transport across the BBB.
2. Delivery systems based upon exosomes that are capable of carrying not only medications but also on a genetic level, the delivery of genetic material to the neuronal tissues within the brain.
3. Stimuli-responsive nanocarrier systems that can release drug contents in response to pH, temperature or enzymatic changes.
4. Gene delivery platforms based upon the use of siRNA, mRNA, or CRISPR technology.

Clinical Applications

Despite many brain targeting strategies currently being tested in preclinical settings, some nanomedicine formulations are already undergoing human testing in the early phases. The goal of the researchers is to produce safe, effective, scalable, and regulatory compliant formulations that ultimately provide a benefit to patients. Technologies such as personalized medicine, AI-assisted design of nanoparticles, and multi-faceted nanocarriers will facilitate the discovery of effective treatments for Nipah encephalitis and other neurodegenerative disorders. In general, brain-targeting drug delivery is an emerging method that has tremendous potential for addressing the issues presented by the blood-brain barrier and improving treatment outcomes^{11,12,16,17}

SAFETY, TOXICITY AND REGULATORY CONSIDERATIONS

Safety Considerations

Brain-targeted drug delivery systems must ensure that therapeutic agents reach the CNS without damaging healthy tissues. The techniques used in delivering agents should not disrupt the BBB by increasing exposure of brain tissue to pathogens or neurotoxins, and should have an acceptable biocompatibility for the delivered agents (e.g., nanoparticles, liposomes, and nanogels). The techniques used should also result in minimal accumulation of drug delivery carriers in organs such as the liver, spleen, and kidneys. The drug delivery system should also provide a controlled manner for the release of the agent to achieve site-specific delivery and reduce the risk of side effects. Finally, the long-term effects of agents administered to the CNS must be evaluated on the functional integrity of the CNS and the health of neurons.^{16,17,20}

Toxicity Considerations

Prior to any clinical use of brain-targeted delivery systems, an evaluation of the potential toxicity of these systems must occur.

1) Nanoparticle-Associated Toxicity

Oxidative stress and inflammation in cells of the CNS.

Cellular injury associated with excessive accumulation of nanomaterials within cells.

Potential neurotoxicity to neurons and glial cells.

2) Drug-Associated Toxicity

High concentrations of drugs can have adverse effects on the CNS.

Off-target distribution may damage organs that were not the intended target of and delivery system.

3) Immunological Toxicity

Activation of the immune system against the delivery vehicle.

Development of allergic or hypersensitivity reactions.

Evaluation Methods

In vitro cytotoxicity assays.

Animal toxicity studies.

Evaluation of pharmacokinetics and biodistribution.

Regulatory Considerations

Regulatory agencies require extensive preclinical testing to verify both the safe and effective use of brain-targeted drug delivery systems. Preclinical testing should include:

Safety pharmacology studies;

Toxicity and biodistribution studies;

Assessment of BBB penetration; and

Assessment of the therapeutic efficacy of the delivery system

FUTURE PROSPECTIVES

The other significant way is in combining targeted delivery systems with new anti-Nipah virus treatment approaches that involve monoclonal antibodies, such as m102.4, broad-spectrum antivirals, RNA therapies, and gene-silencing mechanisms. Future clinical and pre-clinical trials should not only assess the effectiveness of such drugs, but also the pharmacodynamics, distribution pattern, neurotoxicity, and neurological effect of them. In addition, the development of animal models and translational tools will be key for the clinical application of brain-targeted delivery of drugs in case of Nipah encephalitis.

In summary, interdisciplinary collaboration of virologists, neuroscientists, pharmaceutical specialists, nanotechnology experts, and clinicians will be required in order to implement the transition

from laboratory methods of treatment to clinical ones in order to decrease the mortality rate and neurological complications caused by Nipah virus infection.^{11,12,17,20}

CONCLUSION

Nipah virus infections can lead to serious complications that affect the nervous system and have very high mortality rates. There is currently no effective treatment for Nipah encephalitis, but one of the major barriers to successfully treating this disease is that therapeutic agents have difficulty achieving an adequate concentration within the infected brain tissue due to their limited ability to pass through the blood-brain barrier (BBB).

Advances in targeted drug delivery have created new opportunities for overcoming these limitations through the use of nanocarrier systems, receptor-mediated transport systems (e.g., interrupting the blood-brain barrier), intranasal delivery routes, or other innovative approaches to the central nervous system (CNS) targeting of agents.

Recent evidence indicates that CNS-targeted drug delivery systems may offer a number of benefits in addition to increased therapeutic effectiveness; specifically, diabetes medications that have undergone CNS-targeted drug delivery trials show improved bioavailability, decreased systemic side effects, and possibly increased potential for clinical use of newly developed antiviral therapies against Nipah virus infection.

While most of these approaches still reside in the preclinical research stage, the rapid advancement of nanomedicine; molecular targeting; and the research on medications for treating viruses or controlling antiviral activity represents a significant opportunity for developing therapies in the future.

Therefore, it can be concluded that CNS-targeted drug delivery will be an innovative and expanding opportunity for providing management capabilities for Nipah encephalitis; the commitment to accelerating research focused on increasing BBB penetration, target specificity, improving safety, and achieving translational feasibility will determine whether effective treatment options are available for this potentially lethal neurotropic infection and if the global response to future Nipah outbreaks will be strengthened²⁰

REFERENCES

1. World Health Organization. Nipah virus. Geneva: World Health Organization; 2024.
2. Centers for Disease Control and Prevention. Nipah Virus (NiV). Atlanta (GA): Centers for Disease Control and Prevention; 2024.
3. Clayton BA. Nipah virus: transmission of a zoonotic paramyxovirus. *Curr Opin Virol.* 2017; 22: 97 - 104.
4. Lo MK, Rota PA. The emergence of Nipah virus, a highly pathogenic paramyxovirus. *J Clin Virol.* 2008; 43(4): 396 - 400.
5. Ang BSP, Lim TCC, Wang L. Nipah virus infection. *J Clin Microbiol.* 2018; 56(6): e01875 - 17.
6. Sanchez-Lockhart M, et al. Henipavirus pathogenesis and antiviral approaches. *Nat Rev Microbiol.* 2021; 19: 507 - 523.
7. Abbott NJ, Patabendige AAK, Dolman DEM, et al. Structure and function of the blood-brain barrier. *Neurobiol Dis.* 2010; 37(1): 13 - 25.
8. Daneman R, Prat A. The blood-brain barrier. *Cold Spring Harb Perspect Biol.* 2015; 7:a020412.
9. Pardridge WM. Drug transport across the blood-brain barrier. *J Cereb Blood Flow Metab.* 2012; 32(11): 1959 - 1972.
10. Pardridge WM. Receptor-mediated peptide transport through the blood-brain barrier. *Endocr Rev.* 1986; 7(3): 314 - 330.
11. Saraiva C, et al. Nanoparticle-mediated brain drug delivery: overcoming blood-brain barrier to treat neurodegenerative diseases. *J Control Release.* 2016; 235: 34 - 47.
12. Teleanu DM, et al. Nanomaterials for drug delivery to the central nervous system. *Nanomaterials.* 2019; 9(3): 371.

13. Torchilin VP. Recent advances with liposomes as pharmaceutical carriers. *Nat Rev Drug Discov.* 2005; 4(2): 145 - 160.
14. Bozzuto G, Molinari A. Liposomes as nanomedical devices. *Int J Nanomedicine.* 2015; 10: 975 - 999.
15. Allen TM, Cullis PR. Liposomal drug delivery systems: from concept to clinical applications. *Adv Drug Deliv Rev.* 2013; 65(1): 36 - 48.
16. Blanco E, et al. Principles of nanoparticle design for overcoming biological barriers. *Nat Biotechnol.* 2015; 33(9): 941 - 951.
17. Mitragotri S, et al. Overcoming the challenges in administering biopharmaceuticals: formulation and delivery strategies. *Nat Rev Drug Discov.* 2014; 13: 655 - 672.
18. Patel MM, Patel BM. Crossing the blood-brain barrier: recent advances in drug delivery to the brain. *CNS Drugs.* 2017; 31(2): 109 - 133.
19. Banks WA. From blood-brain barrier to blood-brain interface: new opportunities for CNS drug delivery. *Nat Rev Drug Discov.* 2016; 15(4): 275 - 292.
20. European Medicines Agency. Reflection paper on nanotechnology-based medicinal products for human use. Amsterdam: European Medicines Agency; 2020.